

J. Clin. Chem. Clin. Biochem.
Vol. 22, 1984, pp. 473–478

Effect of Magnesium Injection on Foetal Development

By T. Günther, J. Vormann

Institut für Molekularbiologie und Biochemie, Freie Universität Berlin and

H. J. Merker

Institut für Embryonalpharmakologie, Freie Universität Berlin

(Received November 25, 1983/March 27, 1984)

Summary: Pregnant rats were subcutaneously injected twice daily with 1.5 ml 150 mmol/l MgCl_2 beginning at day 5 of gestation. By this treatment development of foetal liver and pancreas was enhanced. In the foetal liver, there was a precocious glycogenolysis combined with a precocious development of the smooth endoplasmic reticulum (ER) and an irregular arrangement of the rough ER. In the pancreas, the development of secretory granules was enhanced.

In alizarin red-stained and cleared specimens, the foetal skeleton was less calcified. The injected Mg^{2+} remained extracellular. Altered hormonal secretion and/or an altered activity of the regulatory enzymes of glycogen metabolism induced by the increased extracellular Mg^{2+} concentration are discussed as possible mechanisms for enhanced foetal liver development.

Wirkung von Magnesium-Injektionen auf die Foetalentwicklung

Zusammenfassung: Schwangeren Ratten wurde zweimal täglich 1,5 ml 150 mmol/l MgCl_2 -Lösung, beginnend vom Tag 5 der Gravidität, subcutan injiziert. Durch diese Behandlung erfolgte in der foetalen Leber eine vorzeitige Glykogenolyse, verbunden mit einer vorzeitigen Entwicklung des glatten endoplasmatischen Retikulums (ER) und einer unregelmäßigen Anordnung des rauhen ER.

Im Pankreas war die Entwicklung von exkretorischen Vesikeln beschleunigt.

Das foetale Skelett war in geringerem Maße calcifiziert.

Das injizierte Mg^{2+} verblieb im Extrazellulärraum.

Als Mechanismus für die beschleunigte foetale Leber-Entwicklung wurden eine geänderte Hormonsekretion und/oder veränderte Aktivitäten der regulatorischen Enzyme des Glykogenstoffwechsels, ausgelöst durch die erhöhte extrazelluläre Mg^{2+} -Konzentration, angenommen.

Introduction

In tocolytic therapy with β -agonists 20.5% of the newborns exhibit hypotrophy according to the chosen score. When, during tocolysis, patients were supplemented with Mg^{2+} (15–20 mmol/day), only 8.8% of the newborns were hypotrophic, i.e. the same frequency of occurrence as in normal pregnancy (8–11%) (1).

When pregnant rats were injected daily with MgSO_4 (150 mg/kg body weight) from day 5 of pregnancy, the foetal livers at term were well developed with morphofunctional features closely resembling those of the adult organs (2).

From these observations, it seems that chronic Mg^{2+} supplementation during pregnancy may enhance

foetal development. We therefore investigated foetal development, to find at which stage of pregnancy it is enhanced by Mg^{2+} , which foetal organs are concerned, and the nature of the biochemical mechanism responsible for the effect.

Materials and Methods

Wistar rats weighing 200 g were used in the experiments. The rats were maintained under a 12-hour light-dark cycle and at a temperature of about 20 °C. Virgin females were mated from 8–10 a.m. and impregnated females were identified by the presence of copulatory plugs. This day was designated as day 0 of gestation. The animals were fed with Altromin (Altromin, Lage, FRG) (Mg^{2+} content: 83 mmol/kg, Ca^{2+} content: 220 mmol/kg) and tap water (Mg^{2+} content: 0.5 mmol/l) ad libitum.

Morphology

Some of the pregnant rats were subcutaneously injected twice per day at 9 a.m. and 4 p.m. with 1.5 ml 150 mmol/l $MgCl_2$. At day 10, 11, 12, 13, 15, 17, 19, 20 and 21 of pregnancy, two Mg^{2+} -injected and one control rat were killed. Up to day 13 the number of somites was calculated. Foetuses from day 10 to 17 were fixed in toto in 30 g/l paraformaldehyde and 30 g/l glutaraldehyde in 0.2 mol/l cacodylate buffer pH 7.2 for electron microscopy and in 35 g/l formaldehyde for light microscopy and alizarin red-staining. From day 19–21, separated tissues were fixed for electron microscopy and histology. For electron microscopy, postfixation was performed in 10 g/l OsO_4 in cacodylate buffer pH 7.2, followed by dehydration in ethanol, embedding in Mikropal, sectioning with Reichert and LKB microtomes, and contrasting in uranyl acetate/lead citrate. Electron microscopy was performed with Zeiss EM 10 and Siemens 101. For histology, slices were stained in haematoxylin-eosin (HE).

For detection of skeletal development, foetuses from day 19 to 21 were fixed in formalin, stained with alizarin red, cleared in NaOH, and stored in glycerol.

Mg^{2+} metabolism

To determine whether $MgCl_2$ injected into the mothers is taken up by foetal tissues, pregnant rats were injected 7 times every 4 hours with 1.5 ml 150 mmol/l $MgCl_2$ beginning from day 18 at 9 a.m. At day 19, the rats were killed at various times after the last injection.

Foetal blood, heart, liver and maternal blood were taken under pentobarbital (Nembutal®) anesthesia (50 mg/kg). For experimental details see l.c. (3).

In a second experiment, after laparotomy in pentobarbital anesthesia, half of the foetuses in each pregnant rat were s.c. injected with 75 μ l 150 mmol/l $MgCl_2$. After two hours, foetal blood, heart, liver, and maternal blood were taken.

Foetal hearts and livers were freeze-dried. For determination of electrolyte content, the freeze-dried tissues were ashed overnight in a Low Temperature Asher (Tracerlab), dissolved in 0.1 mol/l HCl containing 1 g/l La^{3+} . After dilution with H_2O , containing 1 g/l La^{3+} , Mg^{2+} and Ca^{2+} were measured in an atomic absorption spectrophotometer (Perkin-Elmer, model 300). After dilution with 30 mmol/l LiCl, Na^+ and K^+ were measured in a flame photometer (Beckman). All solutions were made with Millipore water. For details see l.c. (3).

Results

Growth rate

When pregnant rats, fed with the normal Mg^{2+} rich food, were injected with $MgCl_2$ from day 5, the number of the somites in the foetuses at day 10, 11, 12 and 13 was unchanged (tab. 1). Also the weight of the foetuses at day 20 and 21 was unaltered (tab. 2). Therefore, growth rate was not affected under our conditions of Mg^{2+} supplementation.

Tab. 1. Number of somites of foetuses from $MgCl_2$ -injected and untreated pregnant rats. Mean \pm SEM, n = number of foetuses.

Day	n	Control	n	$MgCl_2$ -injected
10	8	5–7	16	5–7
11	25	25.4 ± 0.4	18	23.9 ± 0.3
12	9	35.3 ± 0.2	17	35.3 ± 0.1
13	5	44.6 ± 0.4	8	42.1 ± 0.6

Tab. 2. Weight (g) of the foetuses from $MgCl_2$ -injected and untreated pregnant rats. Mean \pm SEM, n = number of foetuses.

Day	n	Control	n	$MgCl_2$ -injected
20	9	3.49 ± 0.08	18	3.40 ± 0.05
21	12	4.60 ± 0.11	14	4.71 ± 0.07

Morphology

Liver

The deposition of glycogen, usually beginning at day 17, was the same in both foetal groups. However, beginning at day 19, the glycogen content was lower in the Mg^{2+} -treated foetuses than in the controls. From day 20 to 21, in the Mg^{2+} -treated foetuses, the glycogen content decreased, whereas in the controls, glycogen increased further. From this result one can conclude that the lower glycogen content in the Mg^{2+} -treated group is caused by glycogenolysis. Parallel with the reduction of glycogen content, there was an increase in the amount of smooth endoplasmic reticulum (ER). The rough ER, normally ordered in parallel was in part irregularly arranged and in contact with mitochondria. Similar behaviour was seen when the biosynthesis of mitochondria was enhanced (4). Cell membranes, mitochondria and nucleus of the hepatocytes were not affected by Mg^{2+} -treatment (figs. 1, 2).

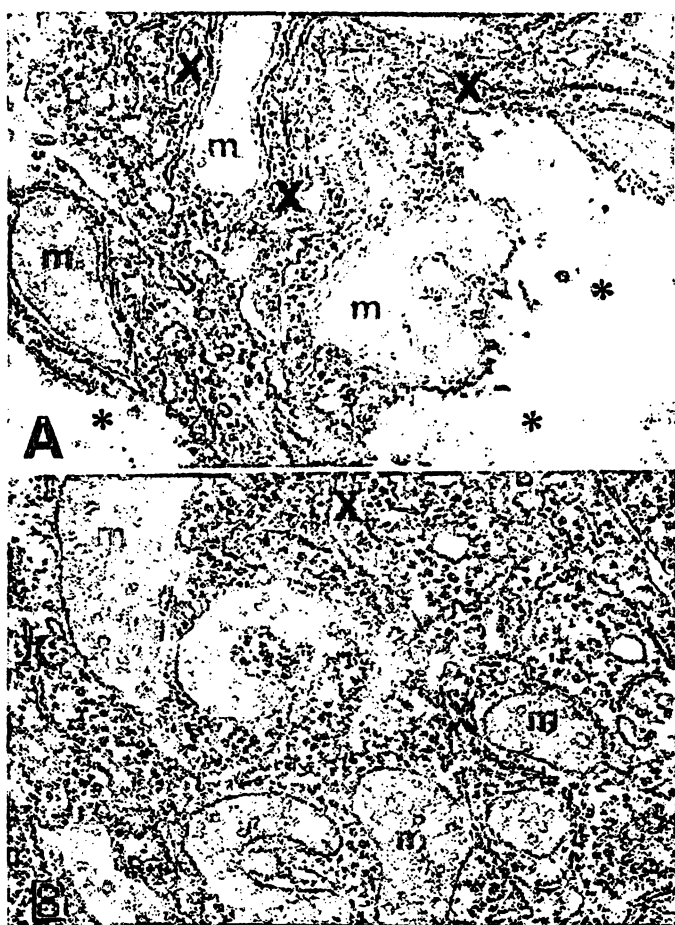


Fig. 1.

- A Part of a liver cell from a control rat foetus day 19 with large glycogen-filled areas (★), mitochondria (m) and cavities of the rough endoplasmic reticulum, packed in a parallel manner (x). $\times 25\,000$.
- B Part of a liver cell from a MgCl_2 -treated rat foetus, day 19. No glycogen deposits and an irregular course of the cavities of the rough ER (x). m = mitochondria. $\times 25\,000$.



Fig. 2. Part of a rat liver cell from a day 20 foetus after treatment with MgCl_2 . Residues of glycogen (★). Increase of smooth endoplasmic reticulum (c). m = mitochondria, N = nucleus. Most of the cavities of the rough endoplasmic reticulum are isolated (x). $\times 25\,000$.

Other cell types of the foetal liver were not significantly changed. The accelerated development by Mg^{2+} -injection did not reach the adult state; adult liver still has less glycogen and more smooth ER than foetal liver after Mg^{2+} -treatment.

Adrenals

As glucocorticoid hormones play a permissive role in glycogen synthesis in foetal liver (5), the development of the adrenals was electronmicroscopically investigated. However, there were no significant qualitative morphological differences in the development of foetal adrenals from untreated and MgCl_2 -treated pregnant rats.

Pancreas

As insulin and glucagon are involved in the regulation of glycogen metabolism and as the ratio insulin/glucagon is perinatally changed (6), the behaviour of pancreatic islets was investigated. The foetal pancreatic islets from MgCl_2 -treated pregnant rats showed no changes in the normal pattern of development, and significant morphological differences between the control fetuses and those from MgCl_2 -treated mothers were not observed.

However, the development of the excretory part of the pancreas was enhanced by MgCl_2 -treatment. Beginning from day 17 and more expressed at later stages, the excretory cells from MgCl_2 -treated fetuses contain more ER and excretory vesicles than those from untreated controls. Some material was already excreted into the ductus (fig. 3).

Lung, kidney

In the lungs and kidneys of fetuses from MgCl_2 -injected and non-injected pregnant rats, no significant morphological alterations could be seen by light and electron microscopy.

Skeleton

In the fetuses of MgCl_2 -injected mothers, the pattern of the skeleton was not altered but the mineralization of all bones was reduced in Alizarin red-stained and cleared specimens. This could be seen best in the tail. As the bones of the tail are the last to be mineralized, the number of tail vertebrae was lower than in the controls (fig. 4).

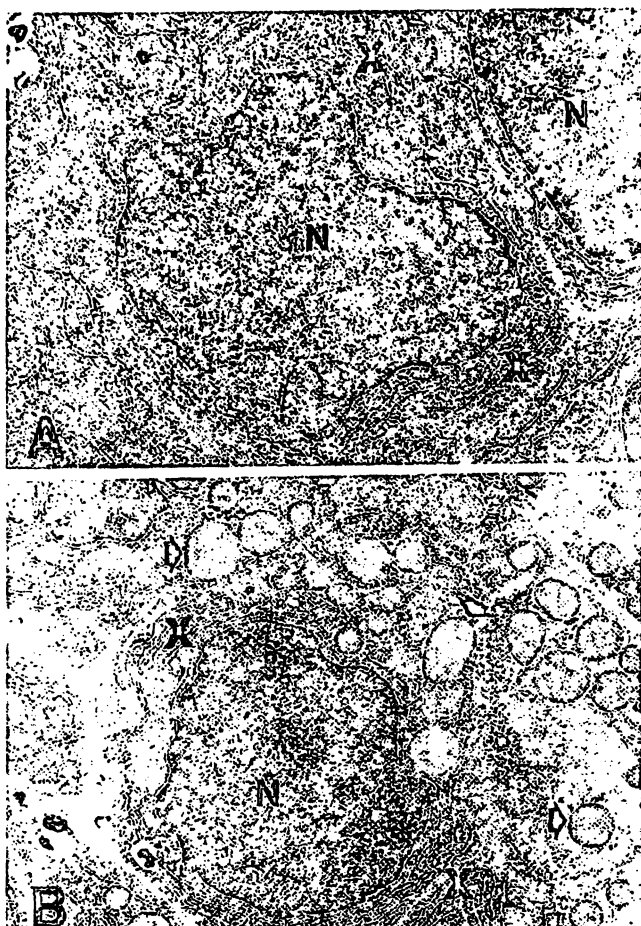


Fig. 3.

- A Exocrine pancreas cell from a control rat foetus, day 19, with numerous cavities of the rough ER (x) and free ribosomes. Note the absence of secretory granules. N = nucleus. $\times 7500$.
- B Exocrine pancreas cell from an MgCl_2 -treated rat foetus with numerous cavities of the rough ER (x) and secretory granules (\diamond). Nucleus (N) with large nucleoli (*). $\times 7500$.

This result is in agreement with in-vitro mineralization experiments. $^{45}\text{Ca}^{2+}$ incorporation in bone was also reduced by competition with Mg^{2+} , when the extracellular Mg^{2+} concentration was increased (7).

Mg^{2+} metabolism

In pregnant rats, subcutaneously injected every 4 hours with 1.5 ml 150 mmol/l MgCl_2 , the serum Mg^{2+} concentration 30 min p.i. was increased to 3.1 mmol/l and dropped rapidly to 1.2 mmol/l within 4 hours after injection (fig. 5). This subsequent fall is due to excretion and to the adsorption of Mg^{2+} to bone.

Serum Mg^{2+} concentration in the corresponding foetal rats only changed between 2.1 and 1.7 mmol/l. Foetal serum Mg^{2+} in this experiment was therefore permanently increased between 10% and 40% for



Fig. 4. The caudal portions of a control (C) and an MgCl_2 -treated (T) rat foetus, day 19, alizarin red-staining and cleared. Note the lower alizarin red uptake (\diamond) or absence (*) of the bone centres in the tail vertebrae.

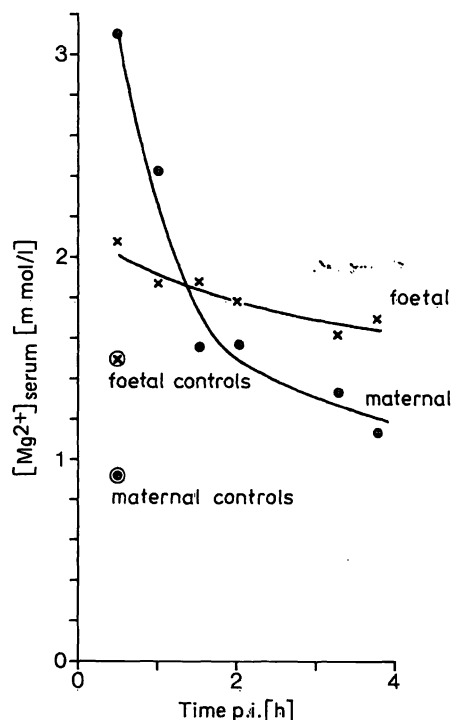


Fig. 5. Foetal (x-x) and maternal (●-●) serum Mg^{2+} concentration at various times after the last s.c. injection of 1.5 ml 150 mmol/l MgCl_2 into pregnant rats (day 19 of gestation). For details see Methods. \otimes = foetal and \odot = maternal serum Mg^{2+} concentration of untreated rats.

24 to 28 hours if compared with uninjected rats. Under these conditions, the Na^+ , K^+ , Ca^{2+} and Mg^{2+} contents in foetal hearts and livers were not significantly changed (tab. 3).

Tab. 3. Mg^{2+} , Ca^{2+} , Na^+ and K^+ content in foetal hearts and livers.

Control: foetuses from non-injected pregnant rats. Mg^{2+} treated foetuses from pregnant rats s. c. injected 7 times from day 18 to 19 with 1.5 ml 150 mmol/l $MgCl_2$. Foetal tissues of each rat were pooled. n = number of pregnant rats in each group. Mean \pm SEM.

	n	Mg^{2+}	Ca^{2+} (mmol/kg dry weight)	Na^+	K^+
<i>Foetal heart</i>					
Control	4	43.5 ± 0.4	5.7 ± 0.1	188 ± 4	490 ± 8
Mg^{2+} -treated	6	43.9 ± 0.3	5.9 ± 0.3	192 ± 5	480 ± 10
<i>Foetal liver</i>					
Control	4	45.2 ± 0.6	2.5 ± 0.1	173 ± 7	532 ± 6
Mg^{2+} -treated	6	45.3 ± 0.4	2.7 ± 0.2	166 ± 13	519 ± 9

Therefore, in a second experiment, foetal serum Mg^{2+} was increased further by direct subcutaneous injection of 75 μ l 150 mmol/l $MgCl_2$ into the foetuses. Two hours p.i. foetal serum Mg^{2+} was increased from 1.38 ± 0.06 mmol/l in the control foetuses to 4.81 ± 0.20 mmol/l in the $MgCl_2$ -injected foetuses. At the same time the tissue Mg^{2+} of $MgCl_2$ -injected fetuses was slightly increased: In the hearts from 43.3 ± 1.4 mmol/kg dry weight to 47.0 ± 0.9 mmol/kg dry weight, and in the livers from 45.7 ± 0.6 mmol/kg dry weight to 50.1 ± 0.4 mmol/kg dry weight. (Values are given as means \pm SEM, n = 6). The other electrolytes were not changed in the foetuses 2 hours after s.c. injection of $MgCl_2$ (not shown). When extracellular and intracellular Mg^{2+} was calculated from the extracellular fluid volume by means of the Na^+ space and serum Mg^{2+} concentrations, no significant increase of intracellular Mg^{2+} in heart and liver was found (not shown). Therefore, the small increase of Mg^{2+} content in the tissues of the $MgCl_2$ -injected foetuses was caused by the increase in extracellular Mg^{2+} concentration.

Discussion

The most significant result of our experiments was the accelerated development of the foetal livers by $MgCl_2$ -treatment of their mothers.

From our morphological results, the decrease of glycogen and the development of smooth ER seem to be combined. As the mechanism for the enhanced development of foetal liver by Mg^{2+} treatment, an increase of the intracellular concentration of Mg^{2+} or other electrolytes can be excluded, because the cellular electrolyte contents were not changed by the

increase in extracellular Mg^{2+} concentration. This also holds for the drastic increase in foetal serum Mg^{2+} concentration by direct s.c. injection into the foetuses. Therefore, the precocious decrease of liver glycogen combined with the precocious development of ER may be produced by other mechanisms. These may be an altered foetal hormone secretion and/or an altered activity of regulatory enzymes of glycogen metabolism.

As the onset and early deposition of glycogen occurs at the normal time point in both groups (day 17), the permissive role of glucocorticoid hormones seems to be unaltered by maternal Mg^{2+} treatment, in agreement with the absence of any morphological alteration of adrenal development.

From a consideration of the biochemical actions of insulin and glucagon, a reduction of the insulin/glucagon ratio could explain the reduction of foetal liver glycogen by glycogenolysis. In normal foetal rats, the insulin/glucagon ratio decreased during the last 2–3 days of gestation (6, 8, 9). However, at the same time, the liver glycogen content increases considerably (5, 8, 9). Therefore, the regulation of glycogen in foetal liver seems to be more complicated, because of the involvement of other components e.g. Ca^{2+} , hormone receptors, adenylate cyclase, cAMP-phosphodiesterase, glycogensynthetase, phosphorylase, phosphorylase phosphatase, synthetase phosphatase (10). Their catalytic activities change during foetal development (8, 9), and their kinetic properties are different from the enzymes in adult liver (8), although they have not been defined in detail.

Thus, the precocious glycogenolysis after Mg^{2+} -treatment, may result from an enhanced reduction of the insulin/glucagon ratio.

An altered extracellular Mg^{2+}/Ca^{2+} -competition at the islet cell membrane is a possible explanation for an altered foetal hormone secretion. When pancreatic islets (from adult animals) were incubated with increased Mg^{2+} -concentration at constant Ca^{2+} -concentration, insulin secretion was decreased (11, 12), although this effect was only significant at rather high Mg^{2+} -concentration (11, 12). However, it is not known whether insulin and glucagon secretion of foetal pancreatic islets shows the same sensitivity to increased extracellular Mg^{2+} as in the adult pancreas.

As glucose-6-phosphatase is bound to ER, the enhanced development of this organelle after Mg^{2+} -treatment may be evidence for increased glycogenolysis by another mechanism. This enzyme, inducible by glucagon (8), can reduce the glucose-6-phosphate

concentration. As glucose-6-phosphate is an allosteric activator of glycogen synthetase, a precocious decrease in glucose-6-phosphate may be responsible for a lower synthesis of glycogen and thus a precocious decrease in glycogen content.

However, other enzymes, regulating synthesis and degradation of glycogen may be also changed by the Mg^{2+} -treatment. Moreover, $MgCl_2$ -treatment may change the concentration of intracellular free Ca^{2+} , which is also involved in glycogen metabolism (10).

Because of the very low cytosolic Ca^{2+} concentration and strong compartmentalization, the participation of Ca^{2+} cannot be determined by measuring the total Ca^{2+} content of the liver. More experiments are needed to clarify the regulatory mechanisms of glycogen metabolism in normal and Mg^{2+} -treated foetal livers. The enhanced development of excretory pancreas may be related to the enhanced development of its ER. The mechanism may be similar to the increase of ER in the foetal liver after Mg^{2+} -treatment.

References

1. Conradt, A. & Weidinger, H. (1982) *Magnesium-Bull.* 4, 103–124.
2. Oliveira-Filho, R. M., Kulay Jr. L., Medeiros, L. O. Medeiros, L. F., Valle, L. B. S. & De Luca, R. (1983) *Gen. Pharmacol.* 14, 291–294.
3. Vormann, J., Förster, R. & Günther, T. (1983) *J. Clin. Chem. Clin. Biochem.* 21, 765–773.
4. Neubert, D., Gregg, C. T., Bass, R., Merker, H. J. (1975) In: *The biochemistry of animal development.* (Weber, R., ed.) pp. 387–464. Acad. Press, New York, San Francisco, London.
5. Greengard, O. & Dewey, H. K. (1970) *Development. Biol.* 21, 452–461.
6. Di Marco, P. N., Chisalberti, A. V., Martin, C. E. & Oliver, T. (1978) *Eur. J. Biochem.* 87, 243–247.
7. Nielsen, S. P. (1973) *Calcif. Tissue Res.* 11, 78–94.
8. Böhme, H. J., Sparmann, G. & Hofmann, E. (1983) *Experientia* 39, 473–483.
9. Margolis, R. N. (1983) *Endocrinology* 113, 893–902.
10. Hems, D. A. & Whitton, P. D. (1980) *Physiol. Rev.* 60, 1–50.
11. Atwater, I., Frankel, B. J., Rojas, E. & Grodsky, G. M. (1983) *Quart. J. Exptl. Physiol.* 68, 233–245.
12. Berggren, O.-P., Bergsten, P., Gylfe, E., Larsson, R. & Hellman, B. (1983) *Am. J. Physiol.* 244, E 541–E 547.

Prof. Dr. T. Günther
Institut für
Molekularbiologie und Biochemie
Arnimallee 22
D-1000 Berlin 33